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# 3D CLEM - combining confocal with FIB-SEM

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MINISTRY OF EDUCATION,  
YOUTH AND SPORTS

**OPEN ACCESS = advanced microscopy for everyone**

**ONLY users' projects and user inspired methodological projects**

## 1. Light microscopy

11 high-end systems

(mostly) **user operated**

## 2. Electron microscopy

FIB-SEM, TEM

+ sample preparation

(mostly) **staff operated**

## 3. Flow cytometry

2 analyzers + 1 sorter

**user and staff operated**

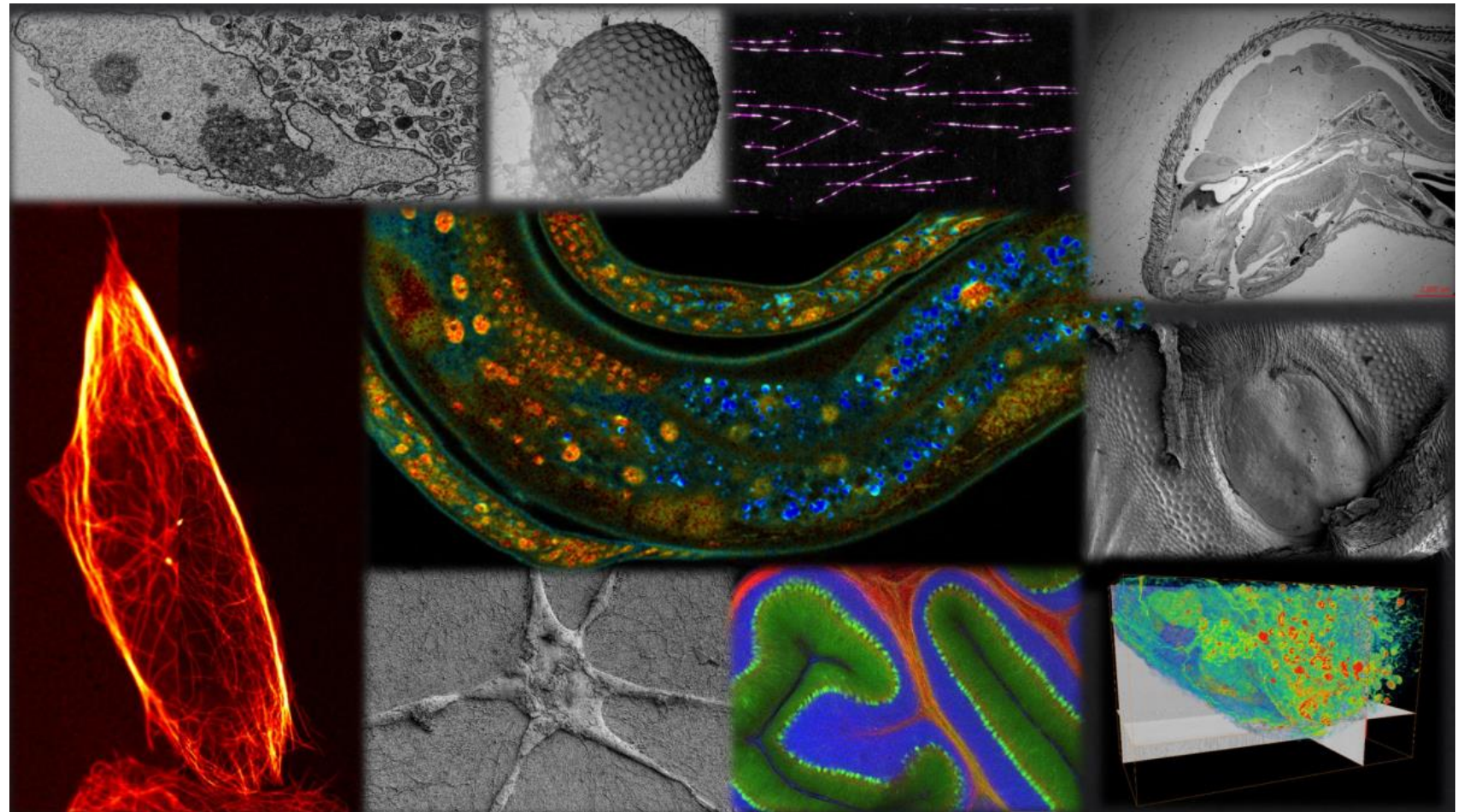
## 4. Image Data Analysis

Commercial software + hardware

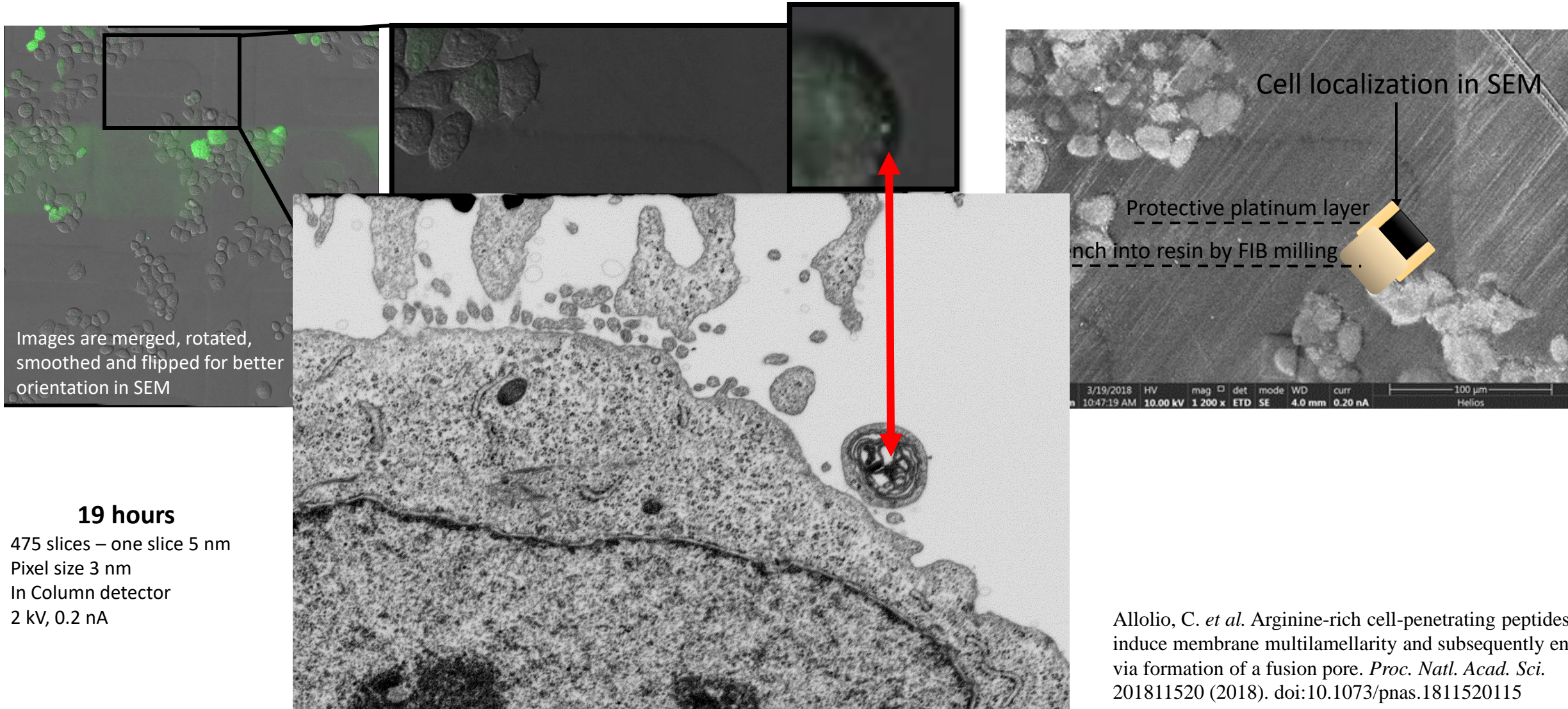
**assistance**

<http://imcf.natur.cuni.cz>

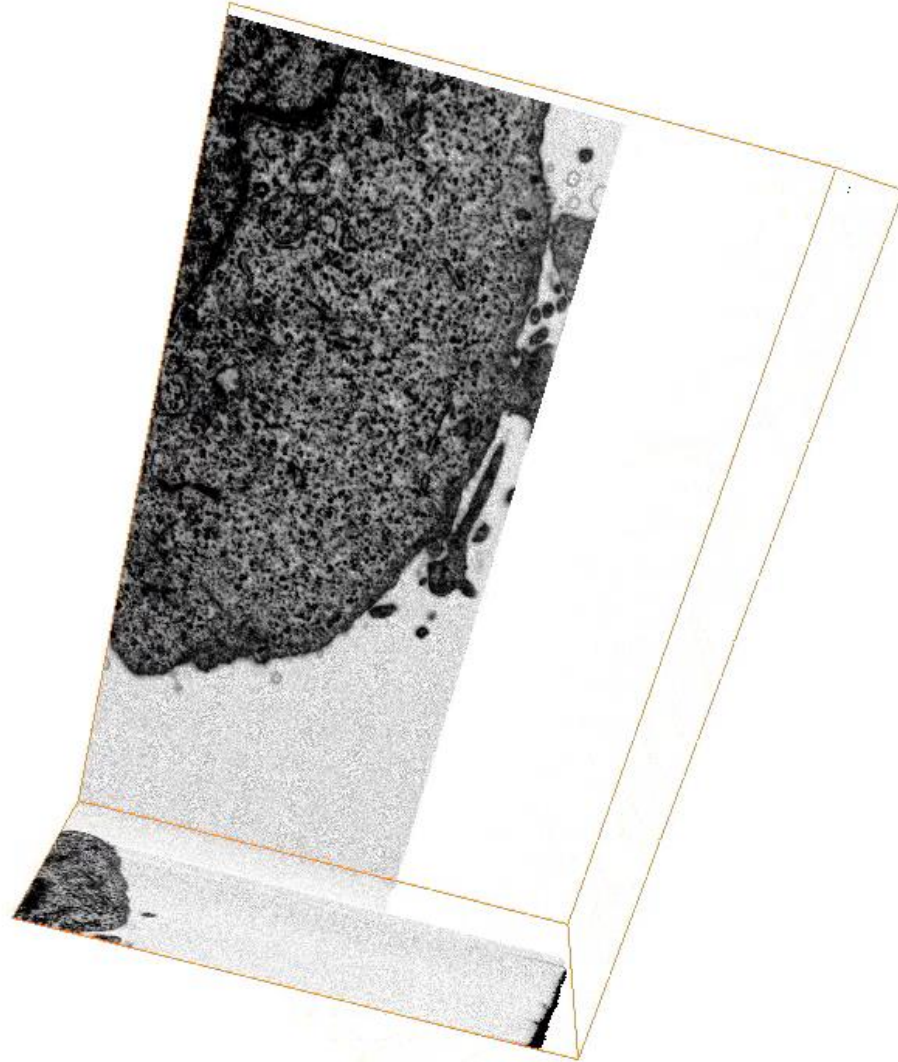
[https://twitter.com/IMCF\\_BIOCEV](https://twitter.com/IMCF_BIOCEV)



# The passive translocation mechanism of arginine-rich cell penetrating peptides



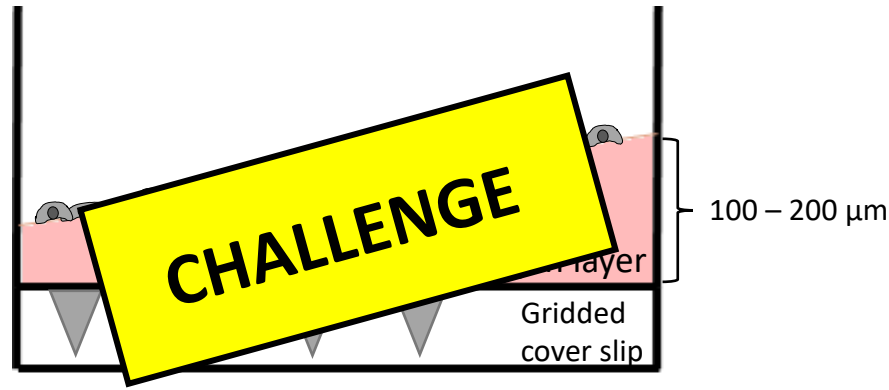
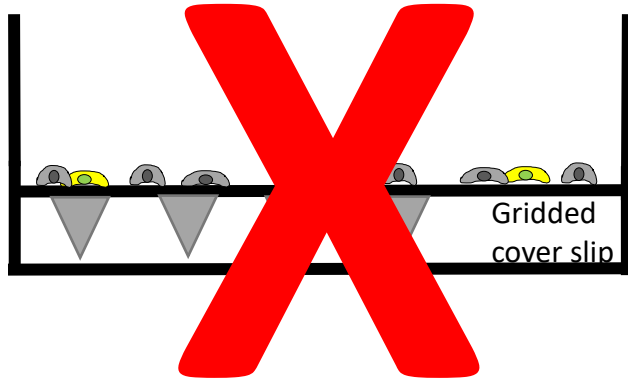
# The passive translocation mechanism of arginine-rich cell penetrating peptides





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# When cells are far away from the cover slip.....



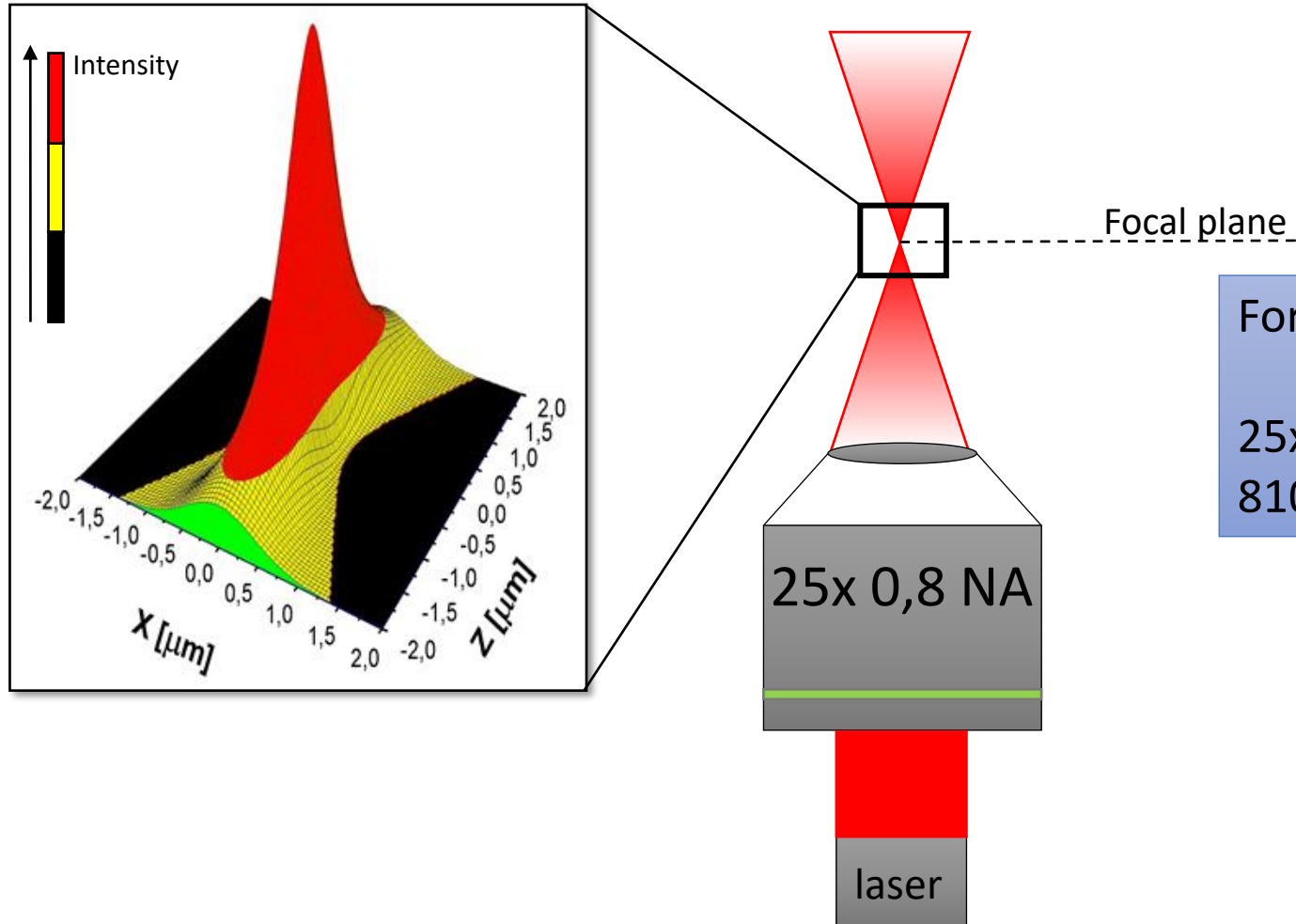
## Limitations

- a cell deep inside the resin can not be localized by an overview SEM
- the depth of milled trench for high quality FIB-SEM is limited (standard depth is 5  $\mu\text{m}$ )
- trimming away the resin up to the targeted cell also removes the imprinted grid ☹️

**The aim** – to analyze cancer cell invadopodia ultrastructure in 3D environment and the transport mechanism of lytic enzymes to ECM degradation sites.

**Method** – CLEM combining fluorescence live cell experiments of cancer cells grown on collagen, expressing genetically labeled matrix metalloproteinases, with FIB-SEM imaging of tumor cells invading the collagen matrix.

# New workflow that extends the pattern from the surface grid deep inside the resin block by using IR laser induced bubbles.

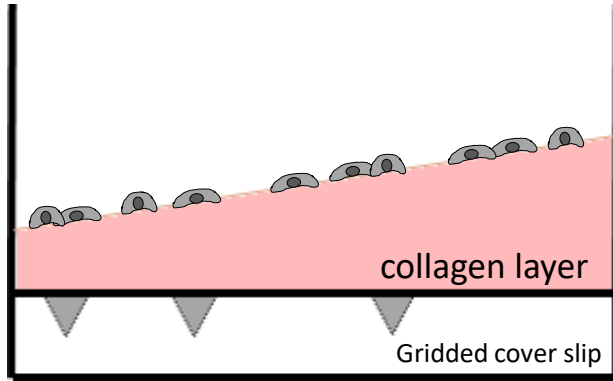


For bubbles burning:

25x WI, 0.8 NA objective

810 nm 2P laser (20-100 mW at sample position)

Cells cultivation on/inside collagen layer  
in a patterned dish



Live-cell imaging (3D confocal  
fluorescent and BF images)



*Carl Zeiss LSM880 NLO*

Sample preparation for FIB-SEM

- Osmium postfixation
- Contrasting with 1% UA
- Dehydration with EtOH
- EPON embedding

Return to confocal  
microscope

**Near-infrared branding (NIRB)**  
IR laser burning bubbles into a resin  
block around the selected cell



*Carl Zeiss LSM880 NLO*

Remove the  
block from the  
patterned dish

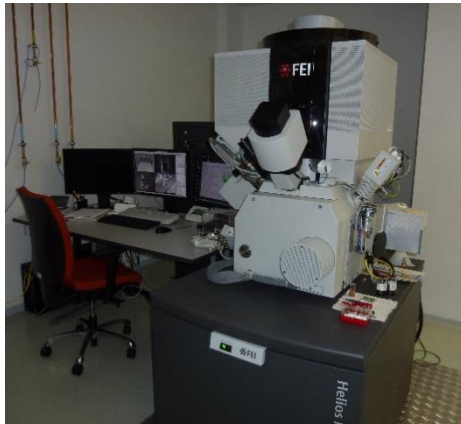
Targeted ultramicrotomy



*Ultramicrotome Leica EM UC7*

Sputter  
coating with  
25 nm Pt

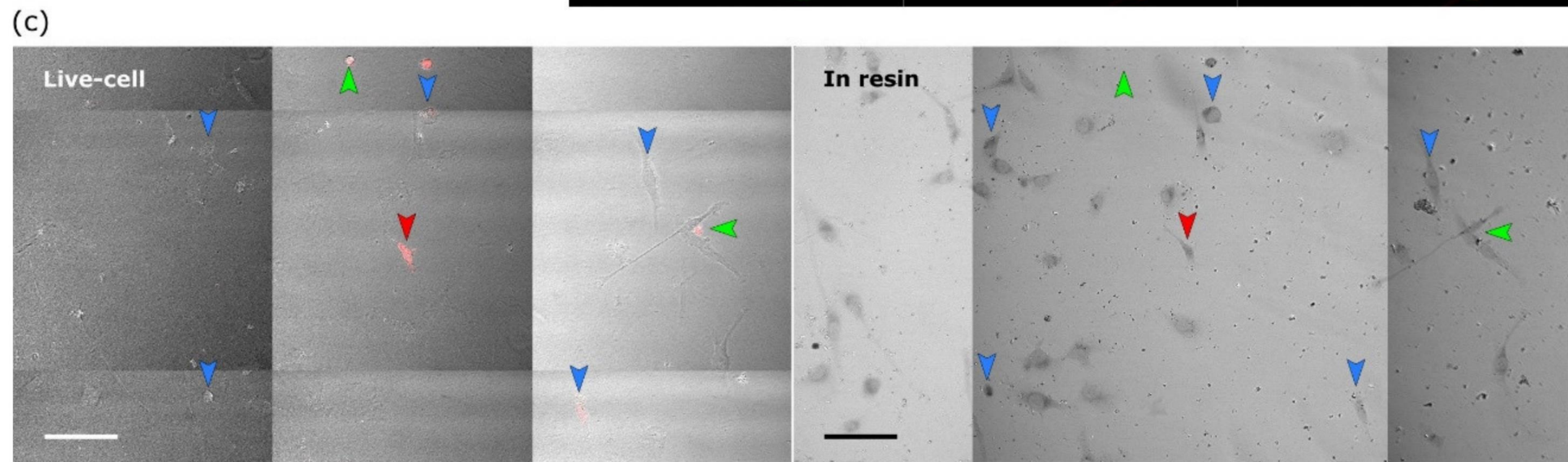
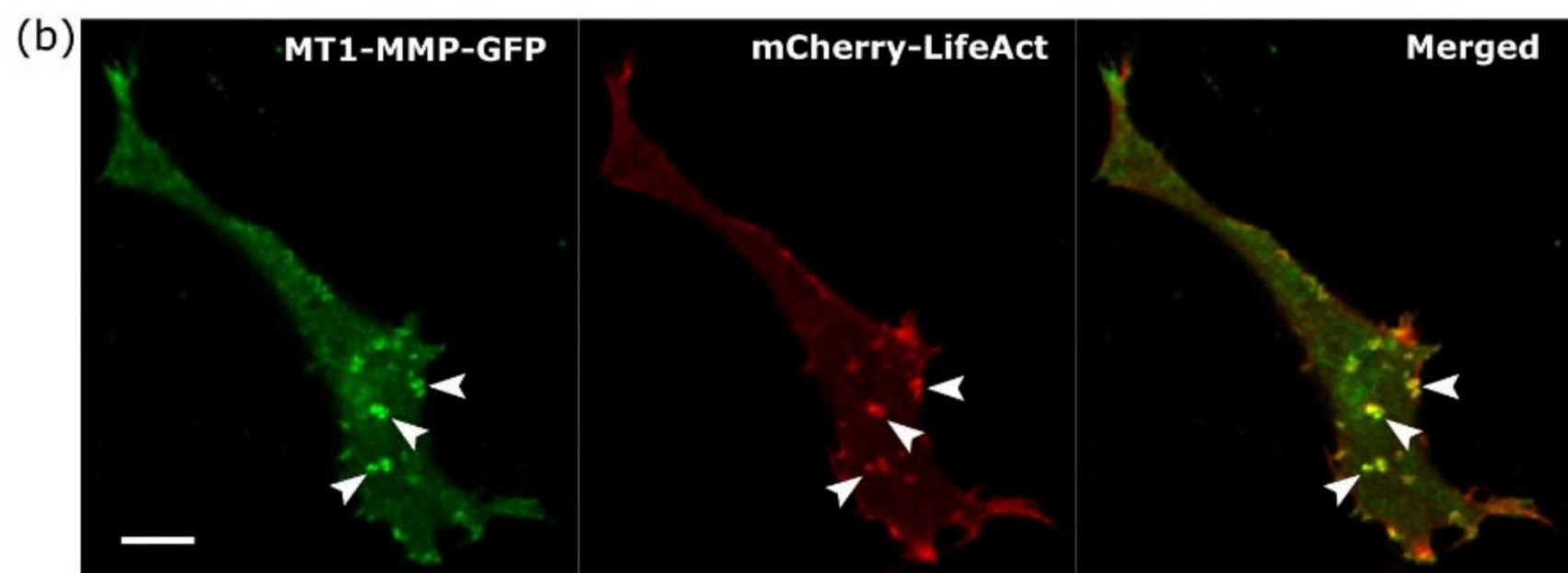
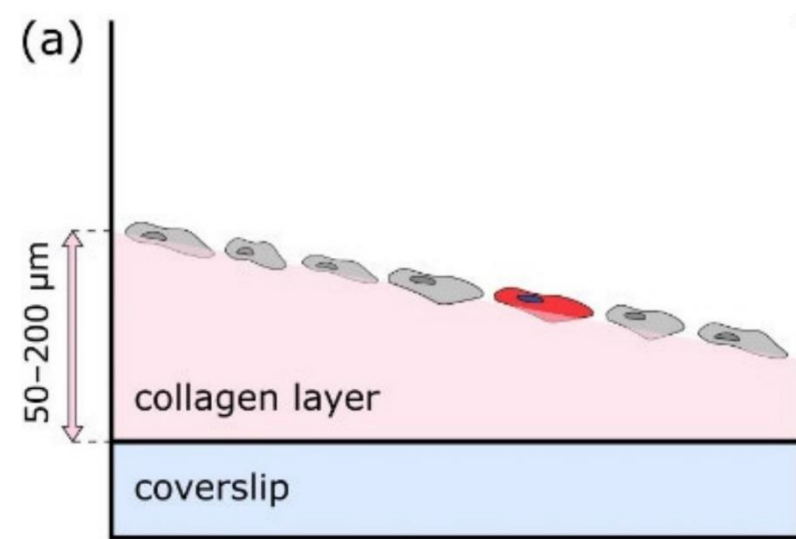
Targeted 3D FIB-SEM



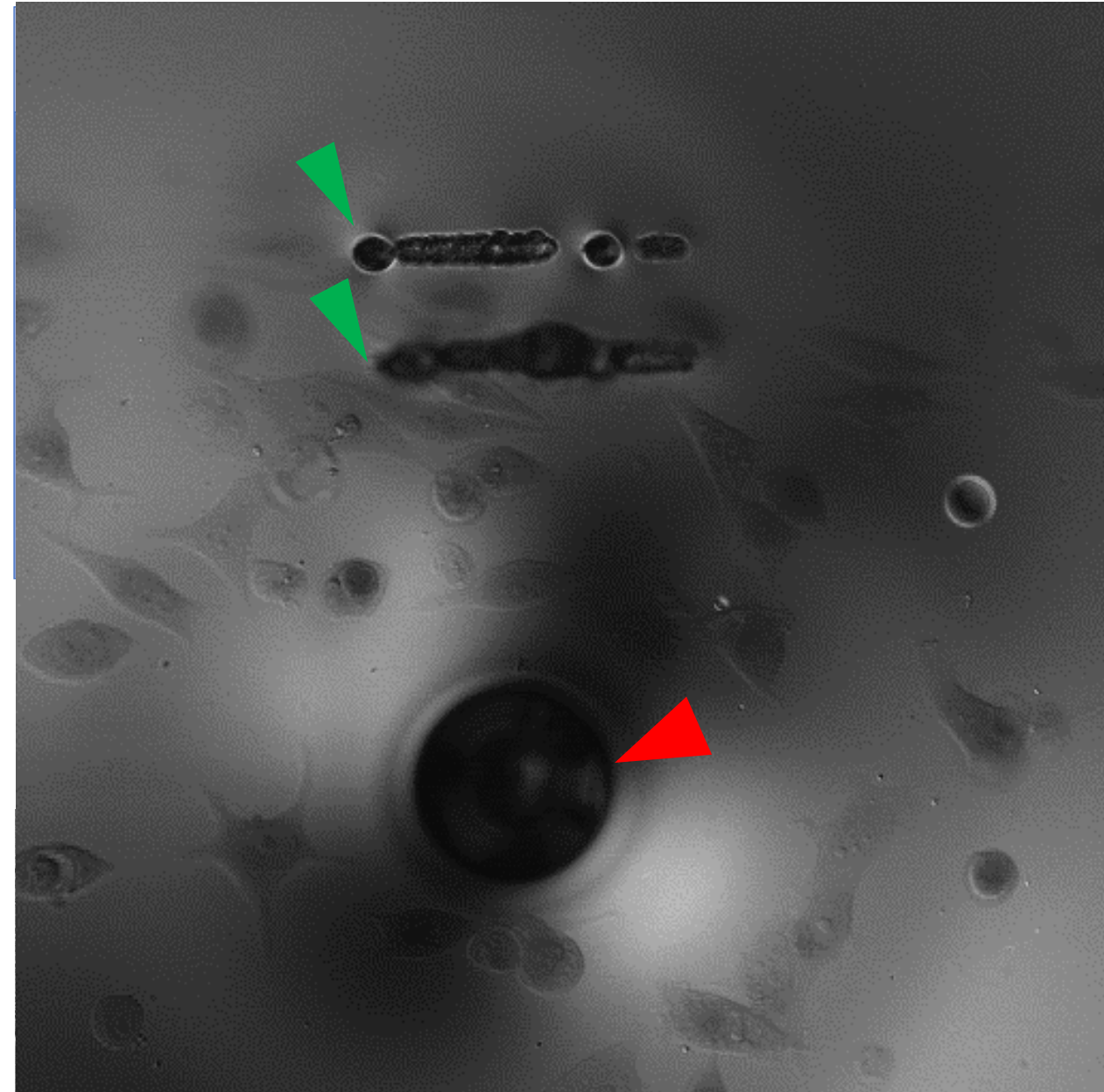
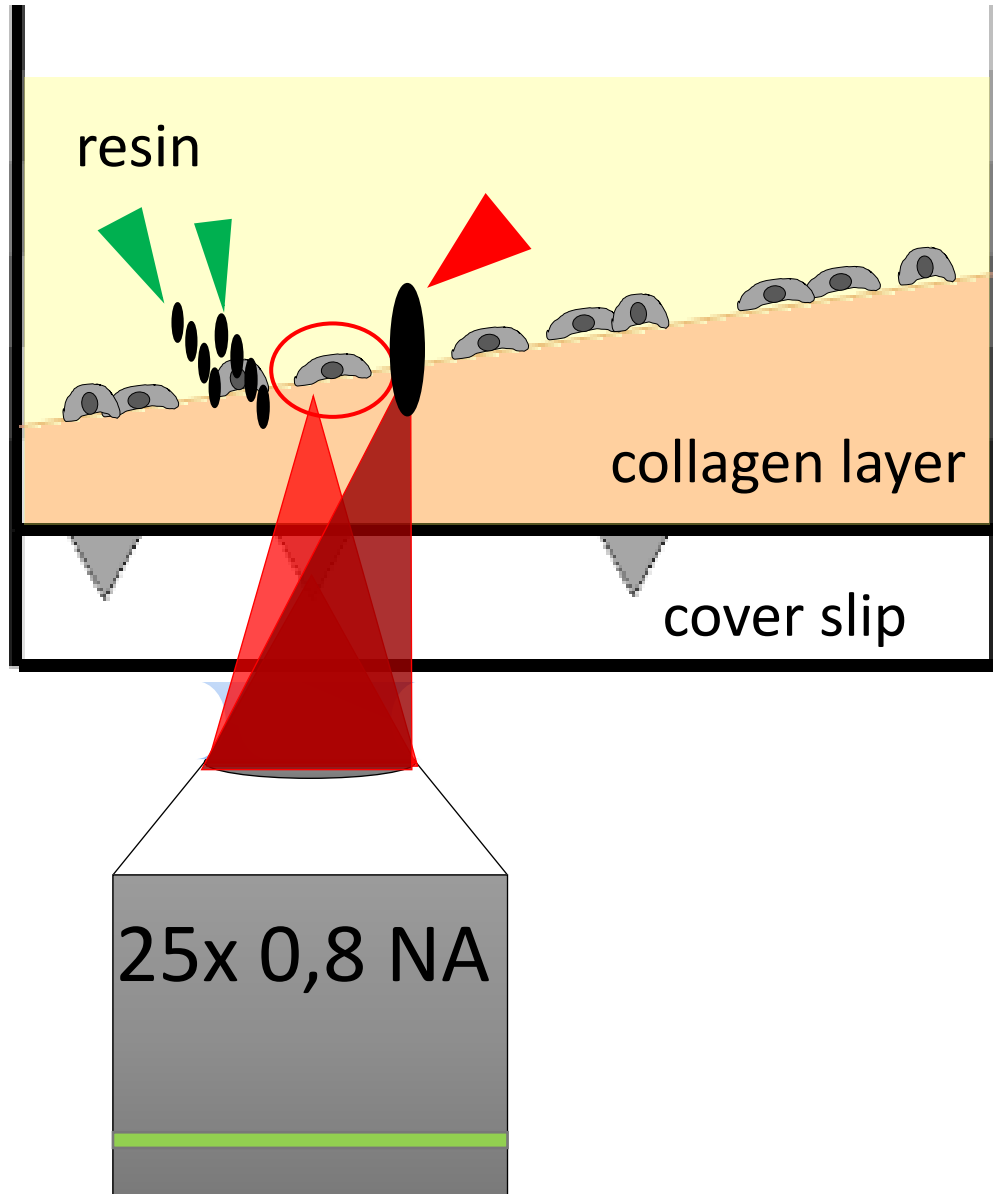
*FIB-SEM FEI Helios NanoLab 660 G3*

Correlation with 3D fluorescent dataset





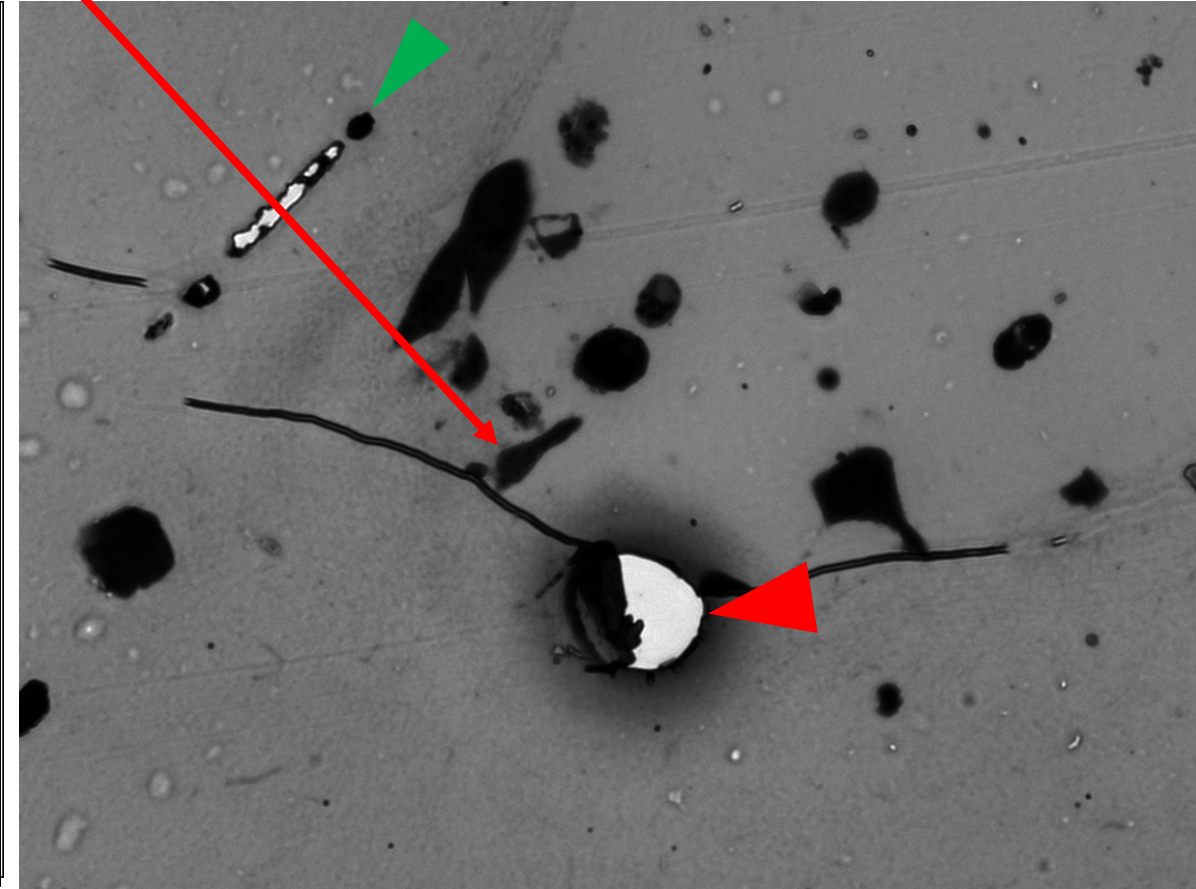
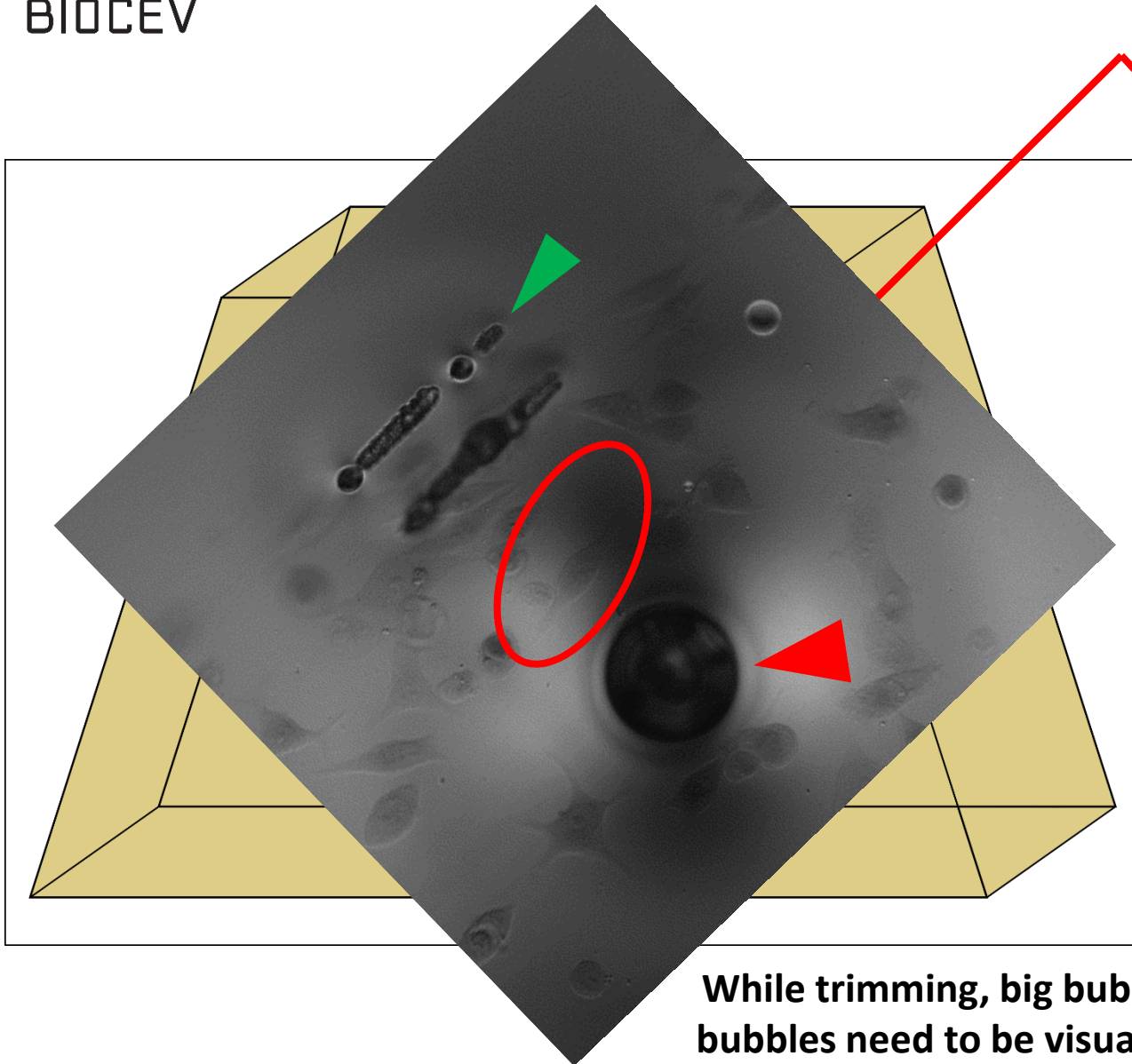
# IR laser burning bubbles into a resin block around the selected cell





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# Bubbles assisted cutting through the collagen layer



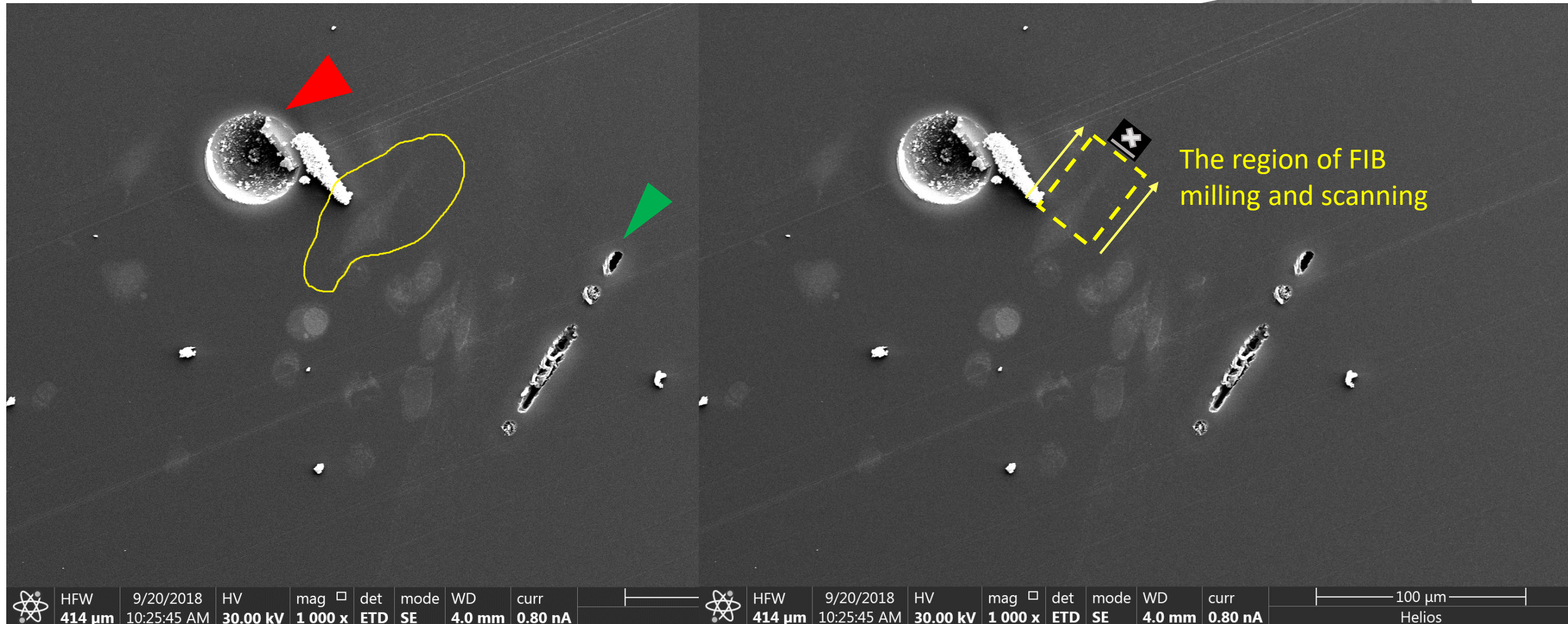
**While trimming, big bubbles are visible with the ultramicrotome optics, cells and line-bubbles need to be visualized by staining sections with toluidine blue.**

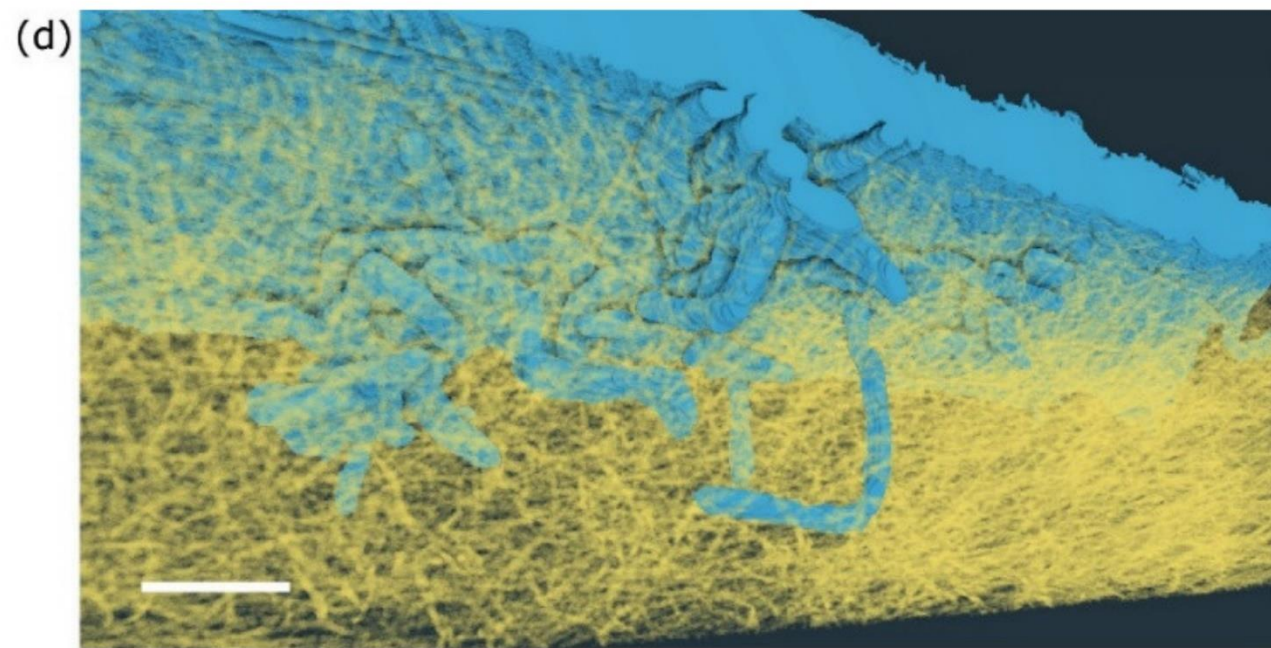
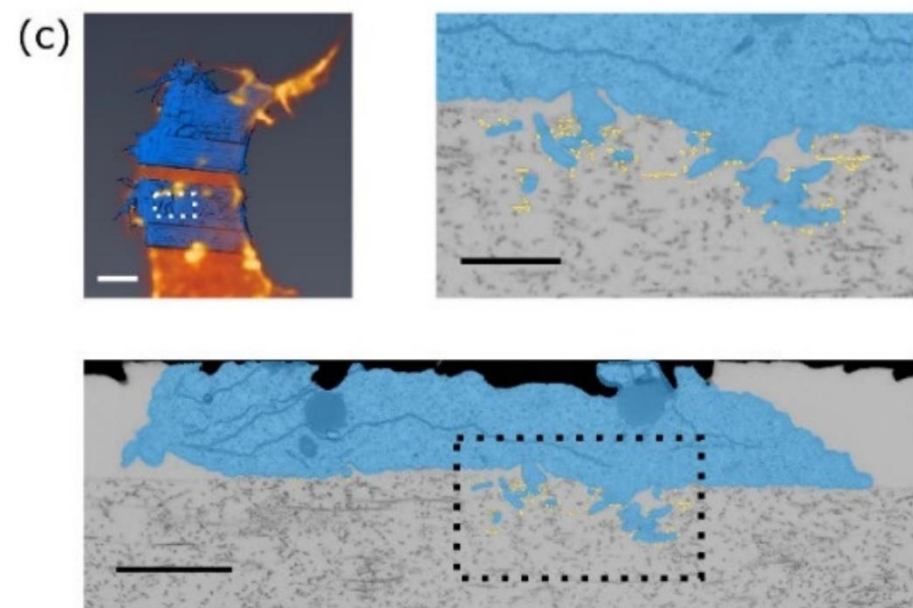
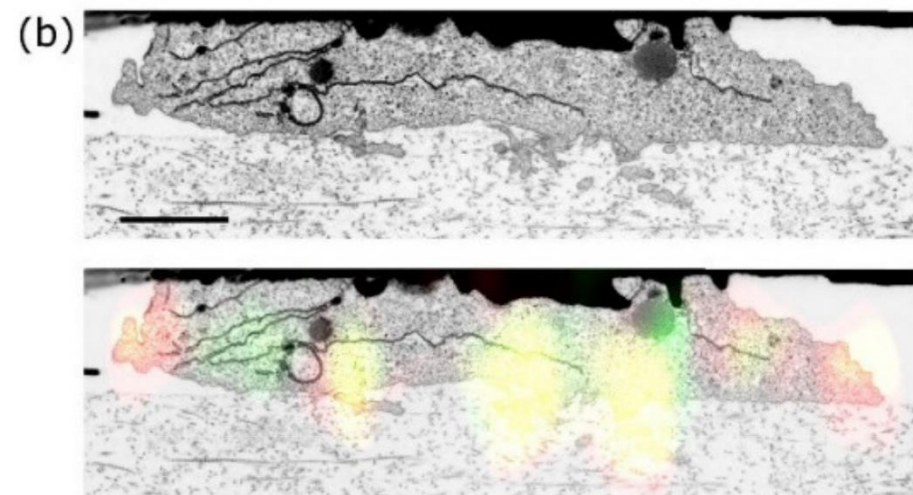
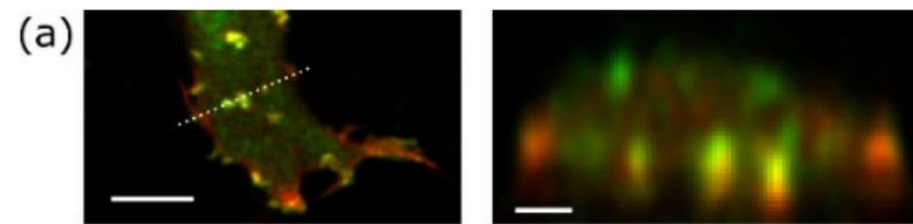


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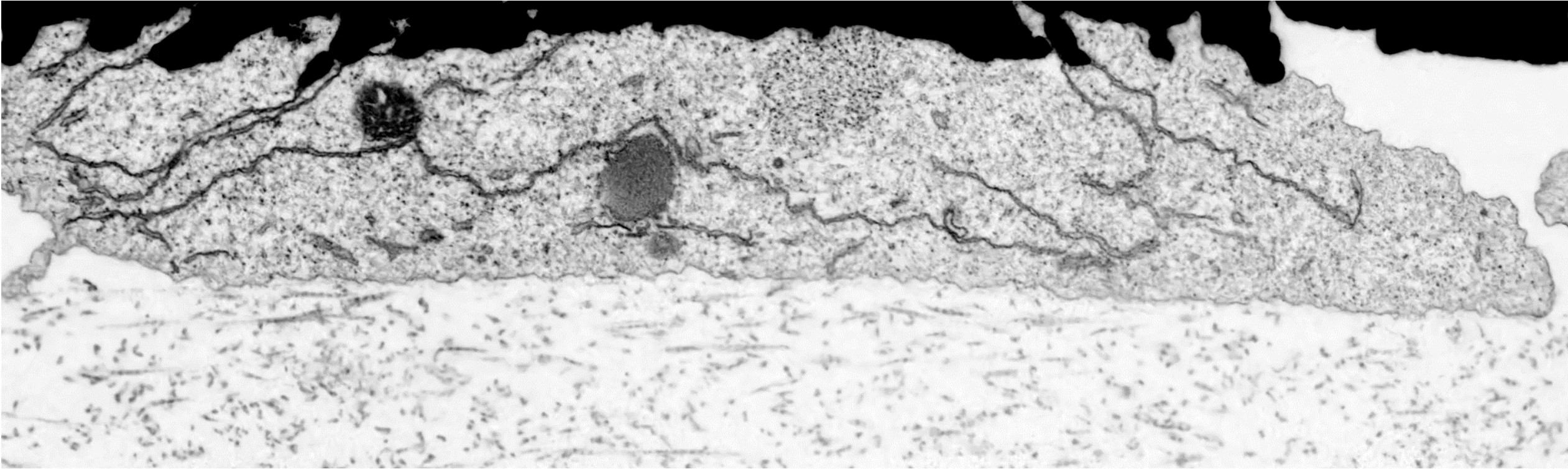
# FIB SEM targeting and milling

After finishing trimming, the block was prepared for imaging by FIB-SEM. The sample was mounted on a regular SEM stub using conductive carbon and then coated with 25 nm of platinum

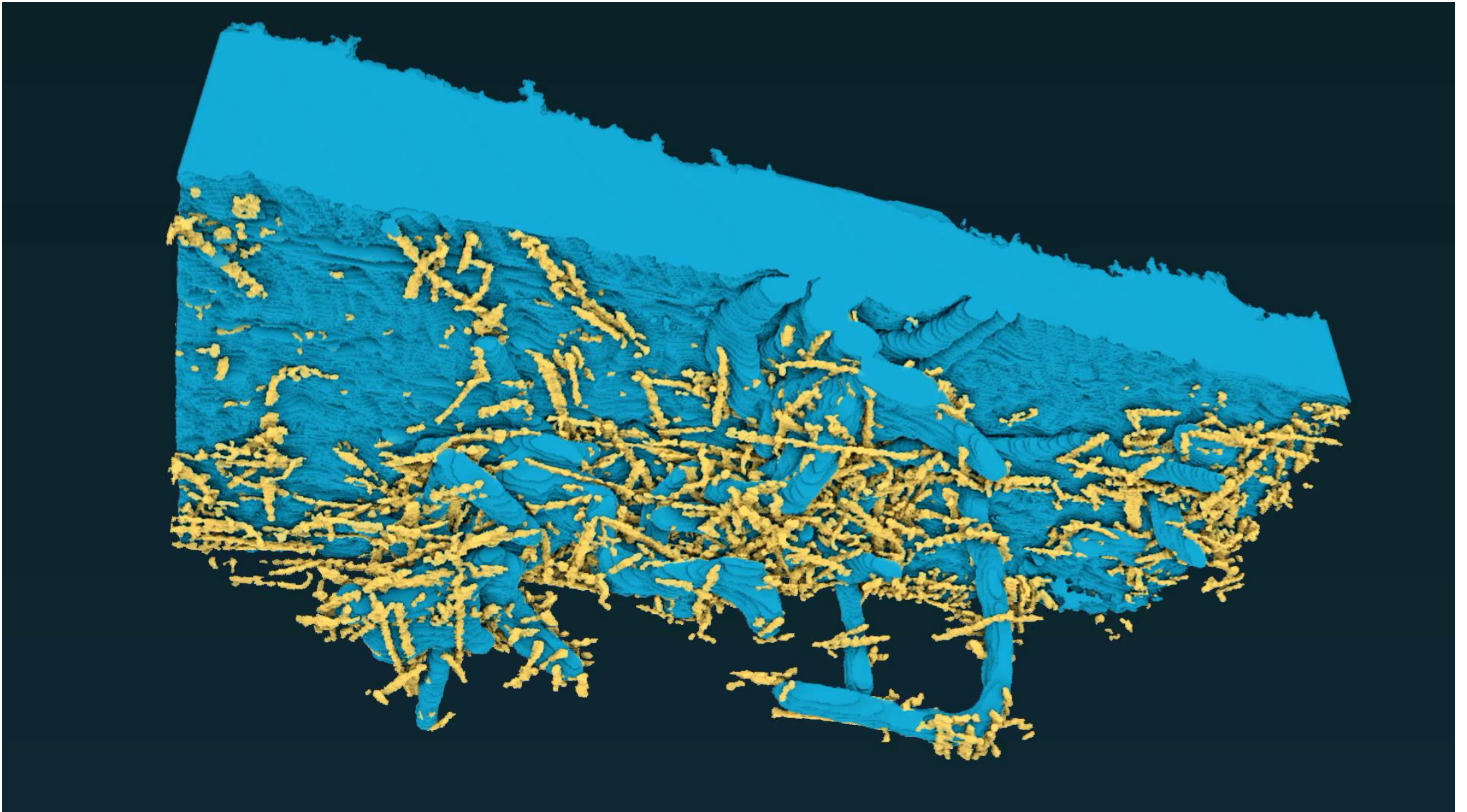




# FIB-SEM movie



# Collagen fibers in a close contact with invadopodia



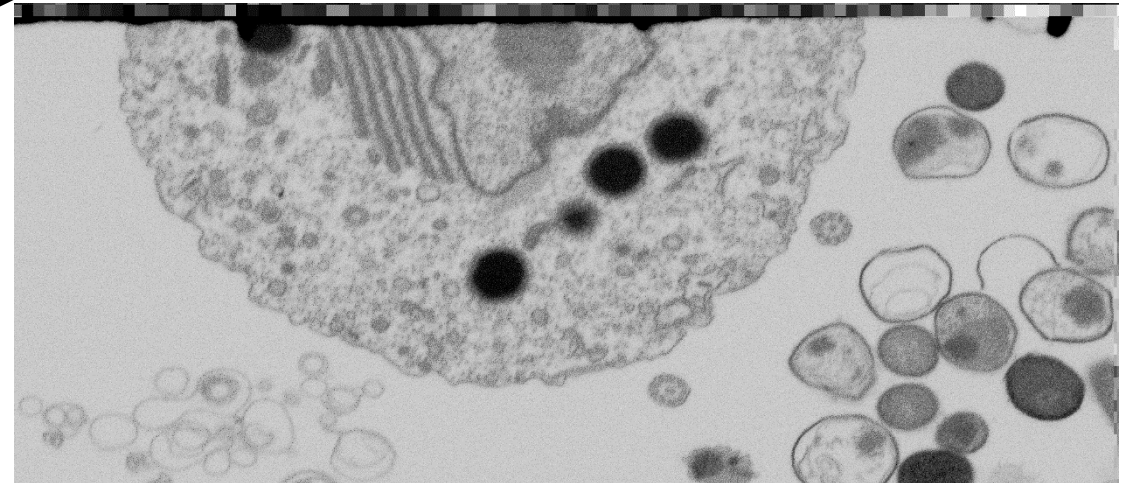
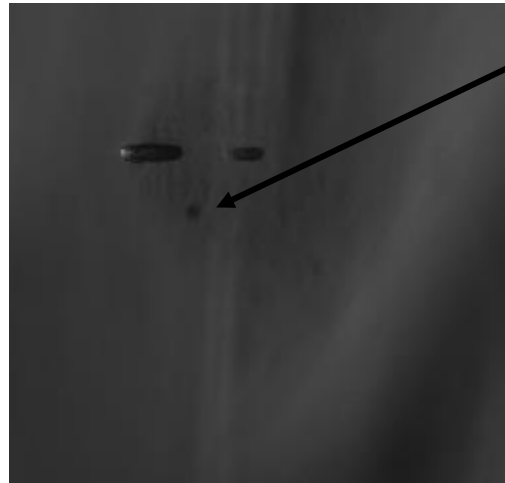
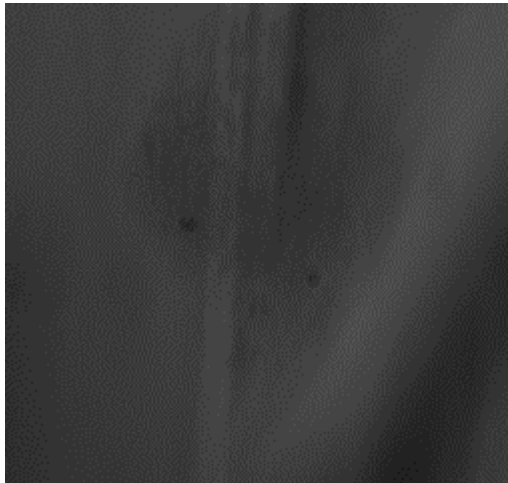


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## Other applications?

- FIB-SEM targeting of samples which are embedded deep in the volume of the resin

Example: a suspension of bacteria and cells of interest



# Acknowledgement

- **ALL our users** (3D-CLEM Ondřej Tolde)
- My IMCF colleagues (<http://imcf.natur.cuni.cz/IMCF/team/>)



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- To MEYS CR (LM2018129) and ERDF (CZ.02.1.01/0.0/0.0/18\_046/0016045)